INSTITUTIONAL BIOSAFETY COMMITTEE
MEETING MINUTES

Vice Chair: M. Maltz-Matyschsyk
Meeting Time: 1:30 PM

February 5, 2026
Web Conference

Attendance

Voting Members	Role / Expertise	Record	Comments
Amalaradjou, Mary Anne	*IBC Chair*	Absent	
Brennan, James	*Animal Care Services*	Present	
Cavallaro, David	*BSO, Secondary NIH Contact*	Present	
Chen, Yupeng	*Biomedical Engineering*	Present	
Chidambaram, Ramaswamy	*Animal Expert*	Present	
Heyduk, Karolina	*Plant Expert*	Present	*Joined at 1:53 pm*
Macierowski, Bobbie	*Local Non-Affiliated Member*	Present	
Maltz-Matyschsyk, Michele	*Vice Chair, Molecular and Cell Biology*	Present	
Miller, Robert	*Local Non-Affiliated Member*	Absent	
Wunder, Elsio	*Pathobiology*	Present	*Joined at 2:00 pm*
Wright, Dennis	*Pharmaceutical Sciences*	Present	
Non-Voting Members & Guests			
Delage, Danielle	*NIH Contact, IBC Program Director*	Present	
Lewchik, Brent	*EHS Director*	Present	
Lucas, Melanie	*RICS Director–Animal Research & Safety*	Present	

The IBC has eleven voting members; a minimum of five members is required for quorum. Seven voting members were present, and quorum was met.

The Vice Chair called the meeting to order at 1:31 PM.

I. Confidentiality Statement & Conflict of Interest

The Vice Chair read the confidentiality statement and reviewed the COI policy with the committee. Committee members were asked to declare potential conflicts and no conflicts were reported.

II. Announcements

1. **Policy & Procedure Updates:** D. Delage provided an update on the ongoing revisions to the IBC Policies and Procedures. To align with the institutional Policy on Policies, all revised IBC policies will be migrated to the new EHS template. The IBC policy statement will be separated from the associated procedures, and policy owners will be clearly designated. The updated policy and procedures documents are expected to be ready for committee review at the next IBC meeting.

2. **NIH Modernizing and Strengthening Oversight of Biosafety:** Listening sessions are scheduled for February 12 and February 26. Updated presentation slides have been posted on the OSP website.

III. Review of Prior Minutes

The Committee reviewed the proposed meeting minutes from January 15, 2026. A motion to approve the minutes as presented was made and seconded. The Committee voted 7 in favor, 0 opposed, and 0 abstentions. The motion carried.

IV. Chair Report:

The Vice Chair provided a report on administrative actions taken since the last meeting.

Submission	PI Name	Committee Action	Status
Renewal #200R2	Senjie Lin	Approved with Conditions on 06/12/2025	Conditions Met – Approval Finalized
Renewal #304R2	Dennis Wright	Approved as Written on 01/15/2026	Approval Finalized

V. Biosafety Officer Report:

Administrative Approvals (those with prior III-D or III-E classifications)

PI Name	Submission	Summary of Request	Approval Date
Erceg, Jelena	81R1A6 Amendment	Adding new transgenic animals. IBC Approved Classifications: III-D, III-F	1/16/2026
Wunder, Elsio	372A2 Amendment	Adding a new cell line and an mRNA vaccine. IBC Approved Classifications: III-D, III-E, III-F	1/16/2026

Laboratory audits are conducted periodically to assess facilities and lab practices to ensure compliance. The BSO presented a summary of recent biosafety audits to the committee.

Audit Report:

- In January, five biosafety audits were completed, three of which had no findings. Corrective actions identified included an out-of-date biological inventory and abandoned samples in a shared freezer showing mold growth on exterior of boxes.

VI. Review of NIH Covered Experiments

Review #1	
PI Name	Jose Manautou
Submission	Renewal #282R2
Title	Cellular Responses to Hepatotoxicants
	Project Overview
Project Description	Acetaminophen overdose accounts for more than half of all cases of chemically induced liver injury. Prior exposure to sublethal doses of acetaminophen or related hepatotoxicants can make hepatocytes more resistant to subsequent toxic insults. The lab aims to understand this adaptive response, focusing on the role of hepatic transporters in detoxification and in the development of tolerance to chemically induced liver injury.
rsNA Nature, Function, & Source Organism	Luciferase reporters – Fireflies, cDNA transcription factors, siRNA targets Mrp4 (drug transporter), MRP4 promoter, FMO3 genes Human, Murine, and synthetic NAs – cDNA (synthetic human and *S. pyogenes* CRISPR/Cas sequences)
Modifications	Knockdowns in rodents via synthetic siRNA lipid nanoparticles or GalNAc conjugates, nonviral, hepatocyte-targeting delivery systems
Experimental Manipulations	The lab previously worked with viral vectors for genetic modification, but is transitioning to LNP and conjugates which should decrease potential risks.
	Risk Assessment & Discussion
Engineering Controls	Preparation of siRNA formulations, lipid nanoparticle suspensions, GalNAc-conjugated materials, and other biologically active reagents will be performed in a certified Class II BSC under BSL-2 containment practices. Tissue processing and handling of animal-derived specimens will be performed in designated laboratory areas using appropriate containment procedures. Intravenous and intraperitoneal injections will be performed in designated procedure areas within the animal facility in accordance with approved animal biosafety practices. Work surfaces will be disinfected before and after procedures using an EPA-approved disinfectant (e.g., 10% bleach solution followed by 70% ethanol). Equipment coming into contact with biological materials will be cleaned and disinfected after use. Spills involving biological or chemically contaminated materials will be managed immediately using appropriate spill kits and disinfectants in accordance with institutional spill response procedures. Biological samples will be securely contained and labeled during storage and transport. All human cell culture materials, animal tissues, recombinant/synthetic nucleic acid waste, and contaminated disposables will be collected as BSL-2 biological waste and autoclaved prior to disposal. Carcasses and bedding from chemically dosed animals will be disposed of as mixed chemical–biological waste per EH&S guidelines. Sharps (e.g., needles, syringes, scalpels) will be discarded immediately after use in approved puncture-resistant sharps containers.

PPE	Lab Coats or Disposable Gowns, Disposable Nitrile Gloves, Safety Glasses, Close-Toed Shoes, Eye protection (safety glasses or face shields as appropriate), and Surgical masks when required *Note: Gloves will be changed frequently, particularly after handling chemically treated animals, recombinant/synthetic nucleic acids, or biological specimens*
Training & Expertise	PI must complete NIH Guideline Refresher Training. Lab members compliant with required trainings. The lab has extensive experience working with vectors to generate transgenic rodent lines as well as for genetic modification of cells. Transitioning to LNP and conjugates should decrease potential risks.
Containment & Special Practices	BSL-2 containment and practices noted as appropriate, with the addition suggested under the Primary Reviewer Comments section.
NIH Classifications	III-E, III-F
Risk Assessment & Primary Review Comments	Overall, this lab presents low to moderate biosafety risk, the work involves human cell lines, recombinant / synthetic siRNA delivered to rodents, all of which require BSL-2 containment. The lab uses appropriate BSL-2 controls, PPE, and waste management will reduce risk.
IBC Motion & Vote	
Committee Discussion	It is requested that the PI address the open notes to clarify comments as outlined in the Open Notes section of the bio-summary.
Motion	A motion was made to approve renewal 282R2, with conditions. The motion was seconded and the committee voted with 7 in favor, 0 opposed, and 0 abstentions. The motion carried.
Committee Action	The committee voted to approve renewal 282R2, with conditions, for a three-year term.

Review #2	
PI Name	John Salamone
Submission	Renewal 305R2
Title	Behavioral Neuroscience of Motivation and Motor Function
Project Overview	
Project Description	The goal of this research is to characterize the role of specific neurotransmitter systems in the regulation of behavior. The lab uses a combination of pharmacological and neurochemical techniques to modify chemical transmission in the brain and study the effect of these manipulations on behavior. The objective is to gain a better understanding the neurochemical processes involved in both normal and pathological behavioral regulation, including a particular emphasis on motor function and motivational processes. Project 1: Rodents are used to investigate the roles that specific drugs have on the brain, mainly compounds that are commonly used to treat neurological and psychiatric disorders. Project 2: This project includes rodent models modified using DREADD methods, also known as chemogenetic methods. A THcre (TH = tyrosine hydroxylase) rodent line will be purchased commercially and an AAV vector will be used to deliver the designer receptor into catecholamine-containing neurons. The designer receptors will be expressed in TH-positive neurons and activated via CNO administration to conduct behavioral experiments. The lab notes that these methods are standard and well-established in the field, although the behavioral techniques used are unique to their laboratory. Project 3: Microdialysis probes will be surgically implanted into the brain to allow fluid to be infused and extracted. Samples containing neurochemicals of interest will be collected, frozen, and later analyzed using high-performance liquid chromatography (HPLC). Project 4: Rodents will be tested on operant touchscreen procedures using liquid and food rewards. They will be assessed for the effects of pharmacological treatments including those commonly used to assess effort-related aspects of motivation in rodent models.
Agent Name	AAV Vectors (purchased through commercial vendor)
Risk Group	RG1
Agent Characteristics	Non-pathogenic viral vector
rsNA Nature, Function, & Source Organisms	Fluorescent reporters – *Discosoma sp.* (sea anemone or disc coral); Inhibit Neuron Receptors – Human
Modifications	AAV and DREADD to express designer receptors in neurons of animal models.
Experimental Manipulations	In-vivo work with wild-type and transgenic rodents; collect and analyze tissue and fluid samples; high performance liquid chromatography. Animals will be perfused with saline followed by formalin prior to extracting brain samples. Tissue samples will be sliced on a microtome, vibratome, or cryostat for histological staining.

Risk Assessment & Discussion	
Engineering Controls	BSC
PPE	Lab Coat, Nitrile Gloves, Safety Glasses, Disposable Gown, Face Shield
Training & Expertise	The lab has extensive experience working with AAV vectors and rodents. PI must complete the NIH Guidelines Refresher Training.
Containment & Special Practices	BSL-1 containment
NIH Classifications	III-D-4, III-E-3, III-F
Risk Assessment & Primary Review Comments	Due to a technical issue, information is missing regarding PPE, decontamination, and waste procedures.
IBC Motion & Vote	
Committee Discussion	Clarification is requested regarding the species of rodents used for the projects. The project descriptions should be updated to include biosafety practices employed and personnel must complete training prior to final approval. It is requested that the PI address the open notes to clarify comments as outlined in the Open Notes section of the bio-summary.
Motion	A motion was made to approve renewal 305R2, with conditions. The motion was seconded. The committee voted with 7 in favor, 0 opposed, and 0 abstentions. The motion carried.
Committee Action	The committee voted to approve renewal 305R2, with conditions, for a three-year term.

1:53 pm – K. Heyduk joined the meeting.

Review #3	
PI Name	Carolyn Teschke
Submission	Renewal 319R2
Title	Function of Seca2 in Export of Proteins; Assembly of bacteriophage P22, Phage Therapy, Understanding Giant Viruses
Project Overview	
Project Description	The Teschke lab studies the folding and assembly of proteins using bacteriophage as a model. Project 1: Studies a secretion pathway; focusing on SecA2 and associated proteins from *Mycobacterium sp.* Proteins are expressed in *E. coli* BL21 and characterized. Precursor proteins and associated extracellular enzymes are analyzed as well. Project 2: Virus assembly experiments are conducted primarily with bacteriophage P22 in *Salmonella enterica* for protein expression. Protein variants will be generated by mutagenesis of plasmids and expressed in *E. coli*. In vitro assays are used to study the effects of mutations on assembly and stability of procapsids. Proteins from phage Sf6 and CUS-3 are also studied. Project 3: Examines the stability of various phages by freeze drying to determine potential uses in phage therapy. Project 4: The lab biochemically characterizes the giant Mimiviruses and determine methods to denature the viruses while also keeping some structures intact. *Acanthamoeba castellanii* is used to grow the Mimiviruses.
Agent Name	*E. coli* BL21*, S. enterica*, Bacteriophage, giant Mimiviruses, *A. castellanii*
Risk Group	RG2 and RG1
Agent Characteristics	None noted
rsNA Nature, Function, & Source Organisms	Structural Proteins from Bacteriophages P22, Sf6, and CUS-3; various structural and assorted precursor proteins from *E. coli* K-12 and *Mycoplasma sp.*
Modifications	*E. coli* K-12 and *Salmonella enterica* LT2 strain contain rsNA constructs for various structural proteins
Experimental Manipulations	The lab grows up to 3 L cultures without shaking and works on the open bench using aseptic techniques.
Risk Assessment & Discussion	
Engineering Controls	Sealed Centrifuge Cups and Rotors
PPE	Lab Coat, Nitrile Gloves, Safety Glasses,
Training & Expertise	This lab has extensive experience working with the materials described and personnel are compliant with all EHS Training requirements.
Containment & Special Practices	BSL-2 containment and practices noted as adequate.
NIH Classifications	III-D-1, III-D-2, III-E, III-F

Risk Assessment & Primary Review Comments	The lab grows up to 3 L cultures without shaking and works on the open bench using aseptic techniques.
IBC Motion & Vote	
Committee Discussion	None
Motion	A motion was made to approve renewal 319R2, as written. The motion was seconded. The committee voted with 8 in favor, 0 opposed, and 0 abstentions. The motion carried.
Committee Action	The committee voted to approve renewal 319R2, as written, for a three-year term.

Review #4	
PI Name	Victoria Robinson
Submission	Renewal 325R2
Title	Structural and Biochemical Characterization of Nucleostemin and the Bacterial Translation Factor BipA
Project Overview	
Project Description	The goals of this lab is to conduct structural and functional studies on GTPase proteins including the bacterial protein BipA. The lab uses various pathogens as well as mammalian GTPase nucleostemin. The projects require large quantities of protein for biophysical and biochemical studies, which is prepared through bacterial expression systems using recombinant nucleic acids. The lab maintains attenuated strains of Enterohemorrhagic *E. coli* (EHEC) and *Salmonella enterica*, which pose a lower risk than the wild type strains.
Agent Name	Non-pathogenic *E. coli*; attenuated strains of EHEC and *S. enterica; Salmonella spp.* SB300A
Risk Group	RG1, RG2
Agent Characteristics	Attenuated virulence (EHEC and *S. enterica*); vaccines or treatments are available
rsNA Nature, Function, & Source Organisms	Translation factors from RG1 organisms *E. coli*, *B. subtilis*, *T. maritima*, *V. fischeri*, as well as RG2 organisms *S. enterica*, *P. aeruginosa*, *S. aureus*, *K. pneumoniae*, and *Toxoplasma gondii;* Transcription factors from RG2 *S. enterica*; Structural / functional proteins from *Drosophila melanogaster*
Modifications	Gene targets are all expressed in RG1 *E. coli* BL21 (DE3). EHEC strain attenuated due to lacking the gene to produce Shiga toxin and *S. enterica* lacks virulence factors.
Experimental Manipulations	Structure and function studies using biochemical and biophysical methods to characterize molecular mechanisms of action for genes of interest. Standard molecular procedures for cloning, expression, and protein purification.

2:00 pm – E. Wunder joined the meeting.

Risk Assessment & Discussion	
Engineering Controls	BSC
PPE	Lab Coat, Nitrile Gloves, Safety Glasses
Training & Expertise	Lab members compliant with require EHS trainings and expertise of the PI and lab personnel is more than adequate for the word conducted.
Containment & Special Practices	BSL-2 containment noted as appropriate. While sharps are used, plastic alternatives (e.g. serological pipettes) are used whenever possible. Sharps are discarded in approved sharps containers, liquid waste is decontaminated with 10% bleach prior to disposal, and solid waste is autoclaved.
NIH Classifications	III-D-1, III-D-2, III-E, III-F
Risk Assessment & Primary Review Comments	None
IBC Motion & Vote	
Committee Discussion	None
Motion	A motion was made to approve renewal 325R2 as written. The motion was seconded, and the committee voted with 9 in favor, 0 opposed, and 0 abstentions. The motion carried.
Committee Action	The committee voted to approve renewal 325R2 as written, for a three-year term.

Review #4

PI Name	Joanne Conover
Submission	Renewal 320R2
Title	Neurogenesis and Brain Repair

Project Overview

Project Description	The Conover lab investigates early brain development, with a focus on stem cells and their functions. Their work includes examining cell-division patterns, generating and evaluating new cell types, and assessing their distribution within brain tissue. Fixed human brain tissue samples are analyzed using immunohistochemistry to evaluate the subventricular zone and brain ventricles for various cell markers and transcription factors. Studies in rodent models are used to inform preliminary analyses of human brain development. A replication-defective avian retrovirus is used to express fluorescent proteins in rodents and observe neurogenesis. The lab also investigates how viral infections of the lateral ventricle may induce hydrocephalus. A heat-inactivated influenza A virus (IAV) is used, with and without luciferase, in collaboration with Dr. Verardi, and will conduct these studies in rodent models. The effects of mutations in genes known to impair cells of the subventricular zone will be studied by expressing recombinant human genes in rodents via in-utero electroporation. Neurogenesis and cell fate will be monitored during development in animals expressing recombinant DNA.
Agent Name	*E. coli* K-12, RCAS viral vectors – ALV and ALSV, heat-inactivated Influenza A Virus (IAV)
Risk Group	RG1, RG2
Agent Characteristics	Non-pathogenic strains; Replication-Defective Viral Vectors; Heat Inactivated Virus
rsNA Nature, Function, & Source Organisms	Fluorescent reporters – *Discosoma sp.* (sea anemone or disc coral), *Aequorea victoria* (jellyfish)
Modifications	Work with non-pathogenic E. coli is used for standard molecular work. Replication-defective RG1 retroviral vectors are propagated in a chicken cell line and used to introduce fluorescent reporters in vivo. The piggyBAC transposon system is used for transient transfection of genes of interest via in-utero electroporation. Plasmid DNA is injected and electroporated into the neural stem cells of the lateral ventricles, which are incorporated into somatic cells over time (i.e., neurons and glial cells).
Experimental Manipulations	In vitro and in vivo methods are used. Tissue samples (e.g. brain tissue) are either isolated from animals or obtained from collaborators, and analyzed using immunohistochemistry techniques. Human samples are fixed and imaged using confocal microscopy. Wild-type and transgenic rodents are either purchased from a commercial vendor, or bred in-house.

Risk Assessment & Discussion	
Engineering Controls	BSC, Sealed Centrifuge Cups and Rotors, and Chemical Fume Hood
PPE	Lab Coat, Nitrile Gloves, Safety Glasses, Close-Toed Shoes, Shoe Covers, Surgical Mask
Training & Expertise	PI must complete the NIH Guidelines Refresher Training. Lab member training compliance is all set. The lab collaborates with a virologist for projects involving Influenza Virus.
Containment & Special Practices	BSL-2 and ABSL-2 containment noted as adequate.
NIH Classifications	III-D-1, III-D-3, III-D-4, III-E-3, III-F
Risk Assessment & Primary Review Comments	
IBC Motion & Vote	
Committee Discussion	Transport of rodents was discussed and clarification is needed on testing and procedures resulting from positive or negative tests.
Motion	A motion was made to approve renewal 320R2, with conditions. The motion was seconded. The committee voted with 9 in favor, 0 opposed, and 0 abstentions. The motion carried.
Committee Action	The committee voted to approve renewal 320R2, with conditions, for a three-year term.

VII. Old Business:

None

VIII. New Business:

None

IX. Items from the Floor:

None

No public comments to record.

Vice Chair made a motion to adjourn, which was seconded. Meeting adjourned at 2:16 PM.

Next Meeting: March 5, 2026 at 2:00 PM